Improved multiparametric scrape loading-dye transfer assay for a high-throughput analysis of intercellular communication

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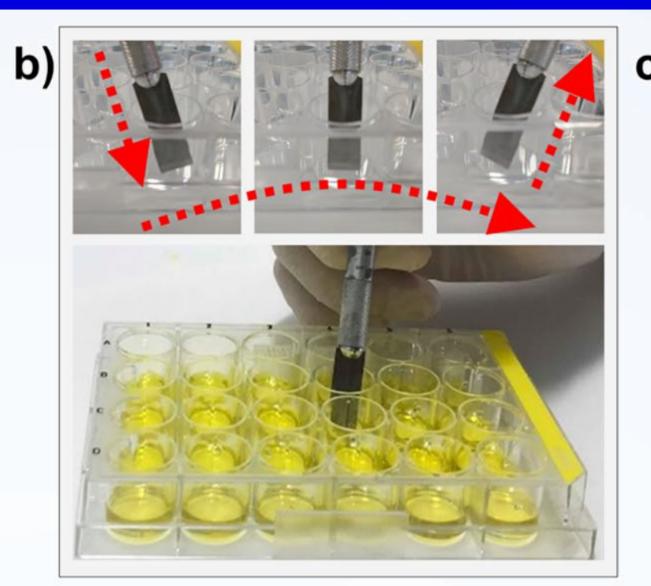
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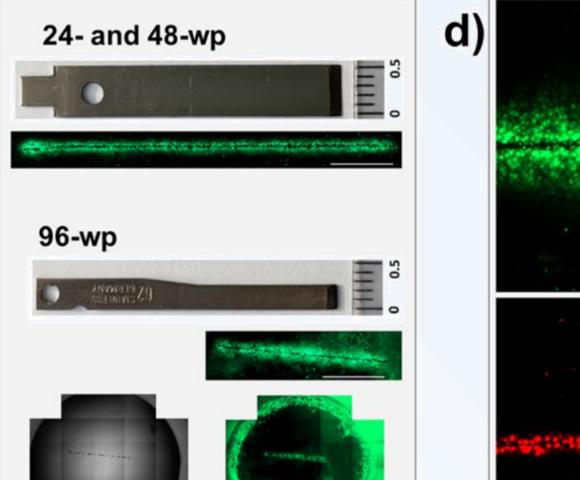
GAP JUNCTIONS

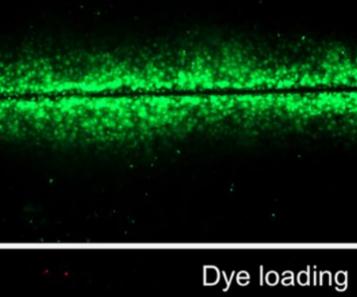
- * Gap Junctions ^[1]
- Closable transmembrane channels
- Connection of adjacent cells
- * Gap Junctional Intercellular Communication^[2]
 - Central homeostatic process
 - Exchange of information, signal molecules etc.

NOVEL MULTIPARAMETRIC METHOD







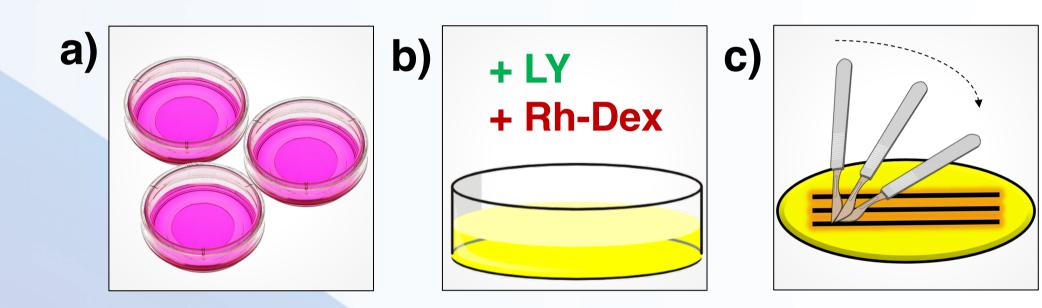


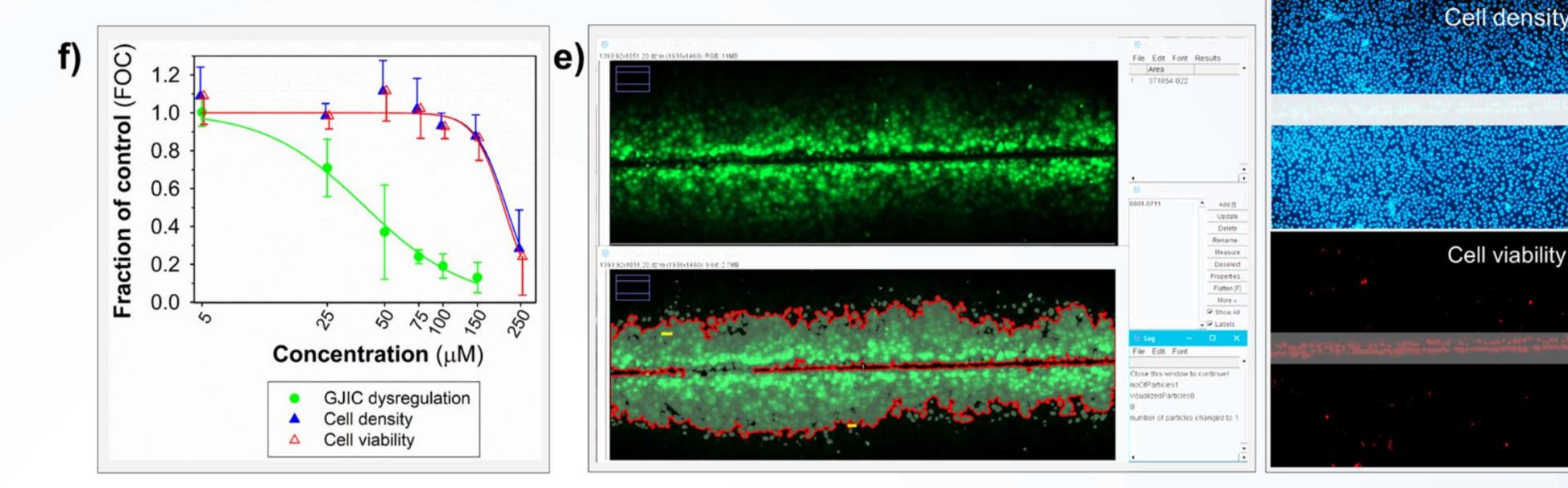
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- Signal transduction, phosphorylation by kinases
- ✤ Inhibition of GJIC^[3]
- Tumour-promoting process
- Dysregulation of apoptosis, contact inhibition, proliferation and various cell functions

TRADITIONAL ASSAY

- Scrape loading-dye transfer (SL-DT) assay^[4]
 - Permeable fluorescent dye Lucifer Yellow (LY) as a tracer of GJIC
 - Impermeable fluorescent dye Rhodamine-Dextran (Rh-Dex) as a "loading control"





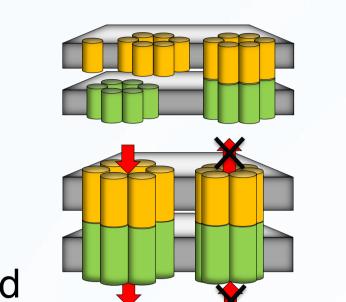
The novel multiparametric *in vitro* assay is suitable for multiple high-throughput-compatible formats, such as 24-well plate (24-wp), 48-well plate (48-wp) and 96-well plate (96-wp; **a**). The evaluation of GJIC dysregulation is based on SL-DT (**b**,**c**) using Lucifer Yellow as a marker of communicating cells (**d-GJIC**) and propidium iodide as a marker of dye-loaded cells along the cut (**d-Dye loading**). The number of total cells is assessed by staining cell nuclei with Hoechst 33342 (**d-Cell density**) and the number of dead cells out of the cut by labelling cells with propidium iodide (**d-Cell viability**). The image analysis and evaluation for all parameters is (semi)automatic using macros in ImageJ software (**e**). Data are expressed as fraction of control (FOC; **f**).

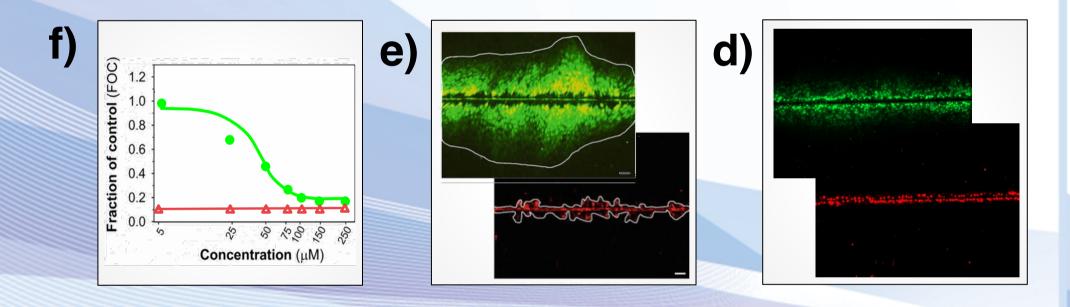
Multiple formats

• 35mm Petri dish, 24-, 48- and 96-well plate ... High-throughput compatible

Multiple endpoints and fluorophores

- Loading control, cell viability
- Gap junctional intercellular communication ...
- ... Propidium iodide
 - ... Lucifer Yellow
 - ... Alternatively Fluorescein 5(6)-isothiocyanate,





Cells are seeded into individual Petri dishes (a), let grow into a full confluency and exposed. The growth (exposure) medium is poured and cells are washed with a buffer, the fluorescent dye mix (Lucifer Yellow and Rhodamine-Dextran) is added (b). After 5-min incubation, three parallel cuts with surgical blade are created (c). Cells are subsequently incubated for a few minutes to allow fluorescent dyes to diffuse through gap junctions. Representative images of dyes' diffusion are taken using fluorescent microscope (d). The area of each of the fluorescent dye is then evaluated using freeware ImageJ software (e). Data are evaluated, compared to negative/solvent control(s) and plotted into graph (f).

* Disadvantages

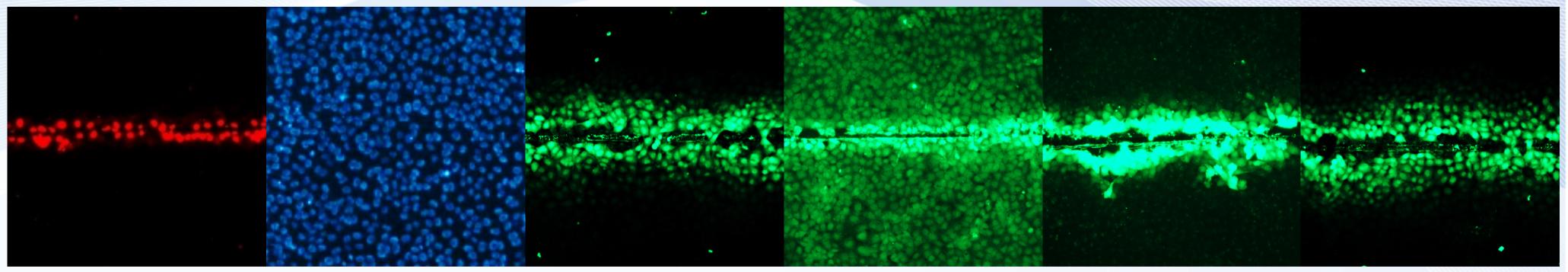
- High material and time consumption
- Costly
 - Fluorescent dyes mix ~ 650 CZK/mL
- Low reusability of materials an chemicals
 - Rh-Dex usable for approx. 1-2 weeks
- Manual performance, not easy to handle
- Only one endpoint
- Manual (subjective) evaluation

- Cell density
- ***** (Semi-)Automatic acquisition and evaluation
- Cost-effectiveness
 - High reusability and stability
 - Lower material consumption

Easy to handle and perform

- Calcein-AM, CF[™] 488A hydrazide
- ... Hoechst 33342
- ... Up to 50-times recyclable fluorescent dyes
- ... Up to 95% less total media consumption
- ... Up to 99% less cells needed for an experiment
- Available at <u>https://go.nature.com/2yplfGT</u>^[5]

Dye loading & Cell viability	Total cell count	Gap junctional intercellular communication			
PROPIDIUM IODIDE (PI)	HOECHST 33342 (HB)	LUCIFER YELLOW (LY)	Fluorescein 5(6)-isothiocyanate	Calcein-AM (C-AM)	CF™ 488A hydrazide (488A)
10 µg/mL, 1500 ms	50 µg/mL, 250 ms	1 mg/mL, 250 ms	(FITC), 1 mg/mL, 500 ms	1 mg/mL, 1500 ms	1 mg/mL, 100 ms
(ex/em 500/640 nm)	(ex/em 350/460 nm)	(ex/em 430/540 nm)	(ex/em 492/518 nm)	(ex/em 496/516 nm)	ex/em 490/515 nm
Filt. RHODAMINE (RHOD)	Filt. DAPI	Filt. ALEXAfluor488 (AF488)	Filt. ALEXAfluor488 (AF488)	Filt. ALEXAfluor488 (AF488)	Filt. ALEXAfluor488 (AF488)

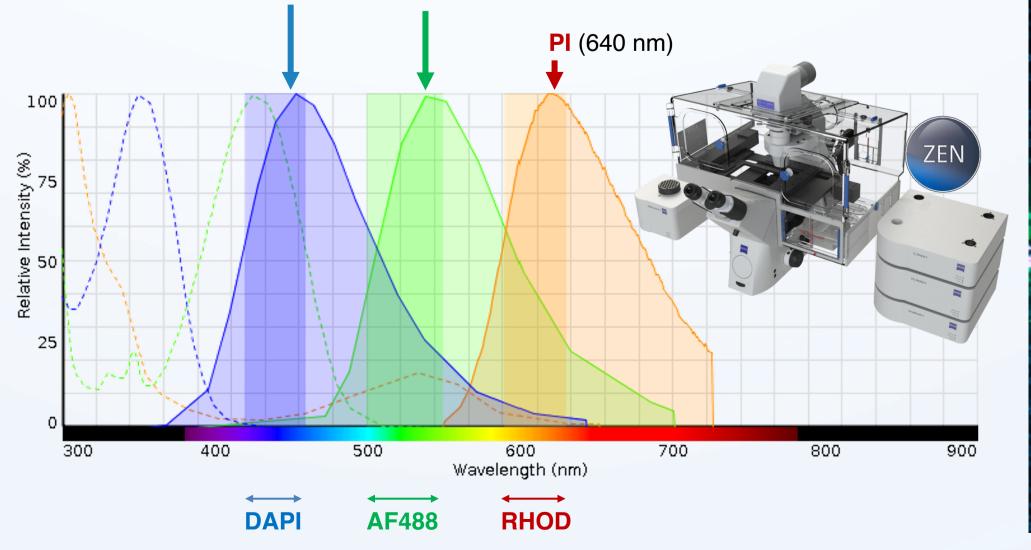


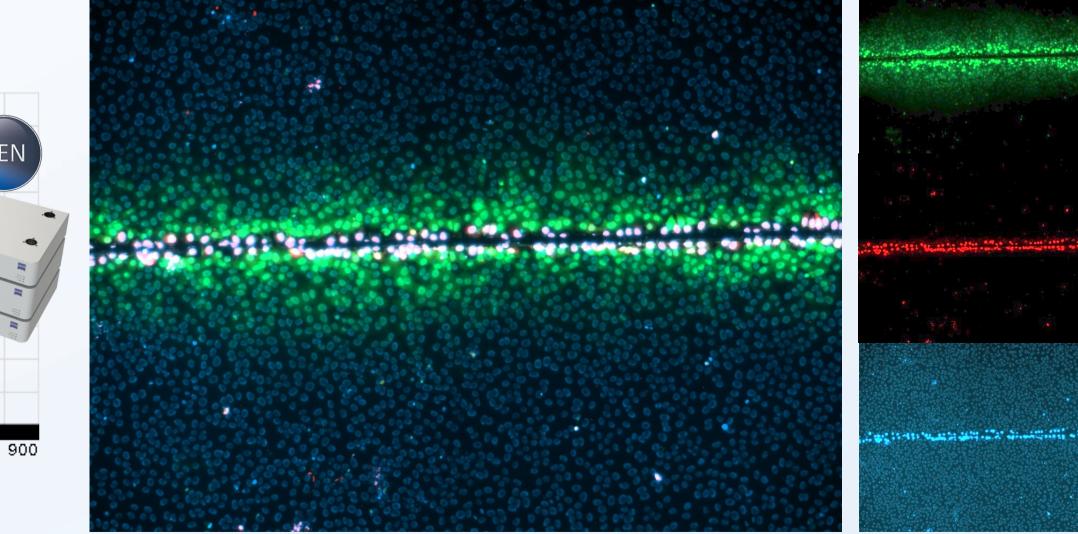
LY (540 nm), FITC (519 nm), HB (460 nm) C-AM (516 nm), 488A (515 nm)

• The evaluator tends to manipulate with the results according to expected values

OBJECTIVES

- * Miniaturisation
- Improved design, high-throughput compatibility
- Better cost-effectivity
 - Lower material consumption
 - Alternative fluorophores
- (Semi)-automatic performance and evaluation
- Automatic acquisition program
- Semi-automatic evaluation in ImageJ
- Multiple endpoints





ACKNOWLEDGEMENT

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