

Toxicokinetic assessment of metabolism-disrupting chemicals in 2D and 3D *in vitro* liver models to improve quantitative *in vitro* to *in vivo* extrapolation (QIVIVE)

OBJECTIVES

- Toxicokinetic approaches using computational modelling
 - Direct toxicokinetics in 2D & 3D *in vitro* models of HepG2 cells
 - Quantitative extrapolation of the *in vitro* and computational results to biologically effective concentrations *in vivo*.
- to determine the effective concentrations of 7 selected metabolism-disrupting chemicals (MDCs)

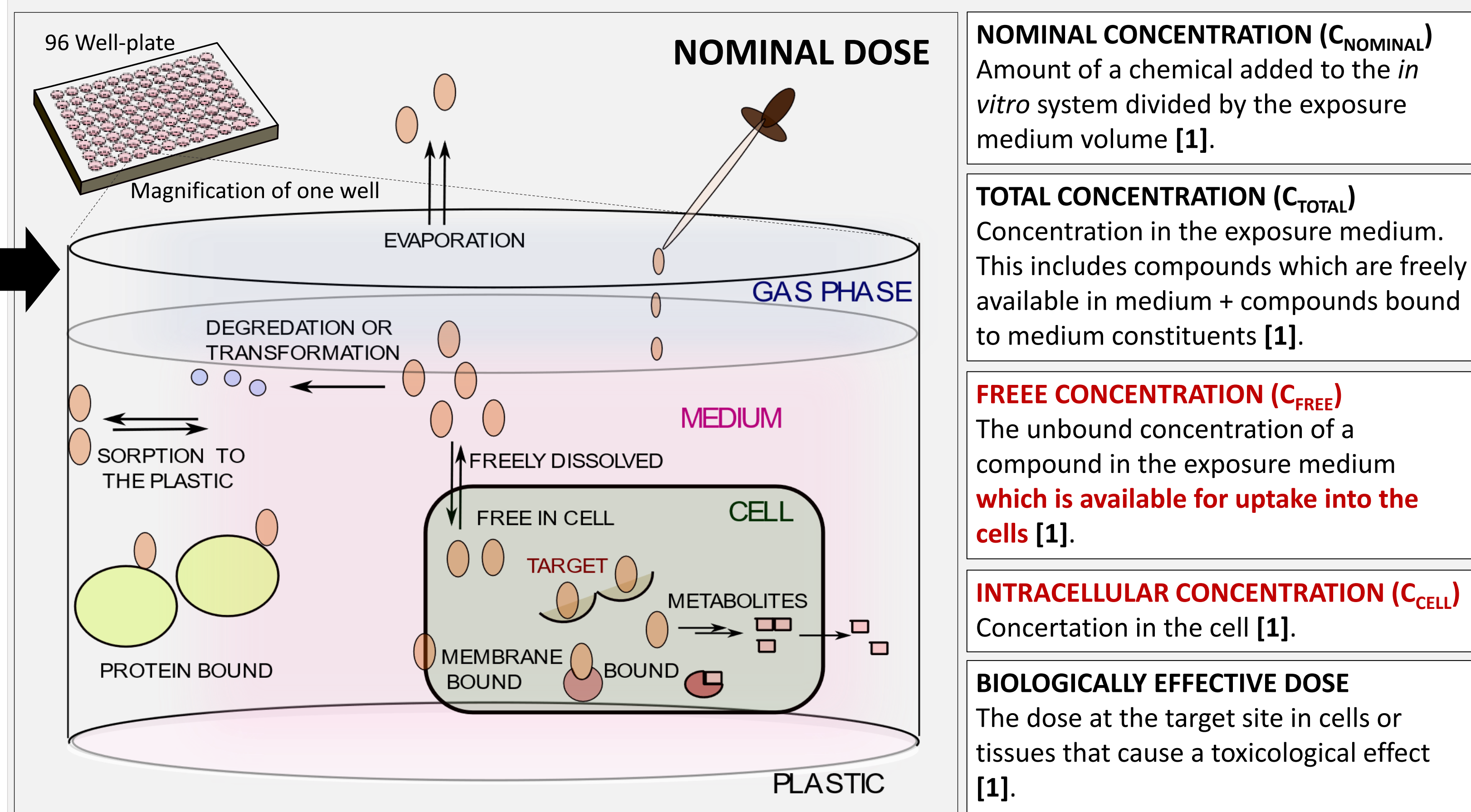
BACKGROUND

What is the problem I need to solve?

- In vitro* human cell-based models play a crucial role in the 21st century toxicology → replacement of animal-based methods; more suitable for high-throughput screening
- So far, toxicity assessment of a tested chemical is mostly based on nominal concentrations (the amount of a chemical added divided by the exposure medium volume).
- However, the added chemical concentration can be affected by several factors!!
- Using the $C_{NOMINAL}$ for risk assessment can lead to FALSE RESULTS!! → UNDERESTIMATION OF THE TOXIC POTENTIAL OF A STUDIED COMPOUND.
- Consequently, target (intracellular), or at least free, *in vitro* concentrations should be used for a precise quantitative *in vitro* to *in vivo* extrapolation (QIVIVE).

Factors which can affect the nominal concentration:

Protein or plastic binding, transformation, metabolism and evaporation of the chemical from the *in vitro* system → **Nominal concentration ≠ effective dose at the target site in cells!**

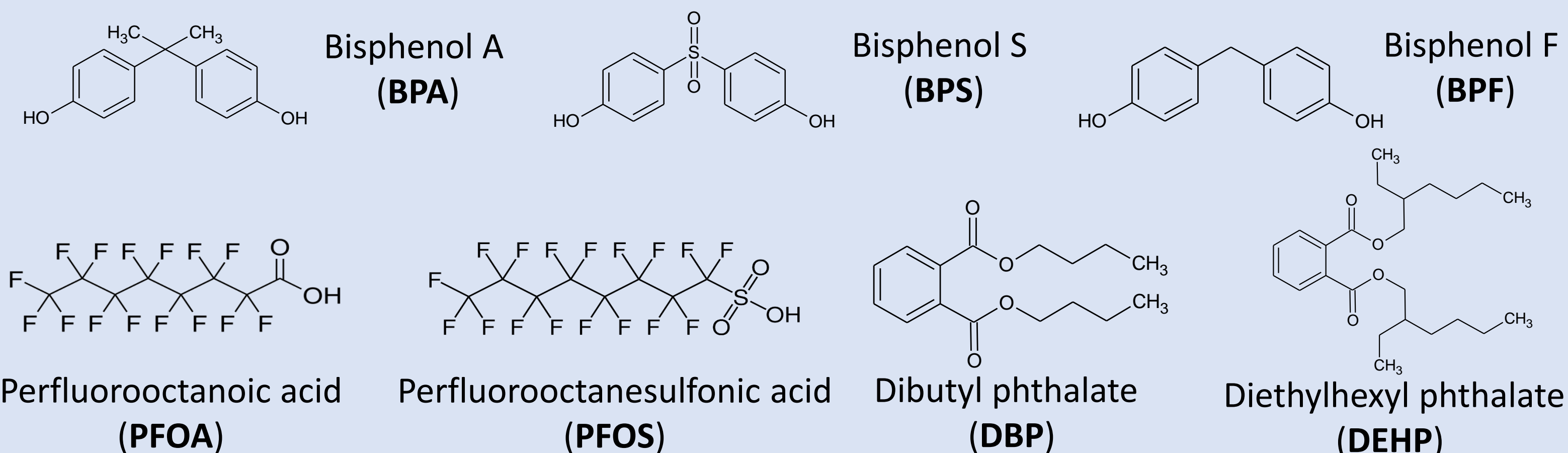


Which chemicals I am going to analyze?

- Endocrine-disrupting chemicals (EDCs) are suspected of causing, beside reproductive abnormalities, metabolic disorders; these EDCs are called metabolism-disrupting chemicals (MDCs).
- MDCs exposures are associated with chronic liver diseases [2].
- Our Cell and Tissue Toxicology Group (SECANTOX) is part of the EU-funded H2020 OBERON project, which aims to establish Integrated Approaches to Testing and Assessment (IATA) to detect EDCs-related metabolic disorders where *in vitro* toxicokinetics and QIVIVE should play an indispensable role.



In vitro toxicokinetics research is being conducted for 7 MDCs from the OBERON project.



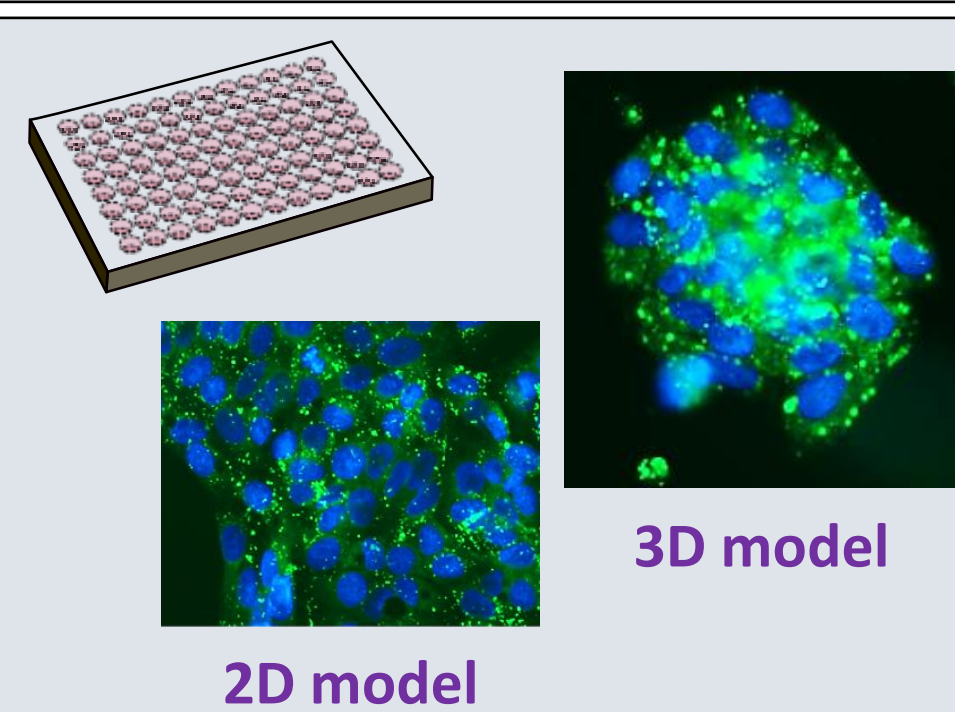
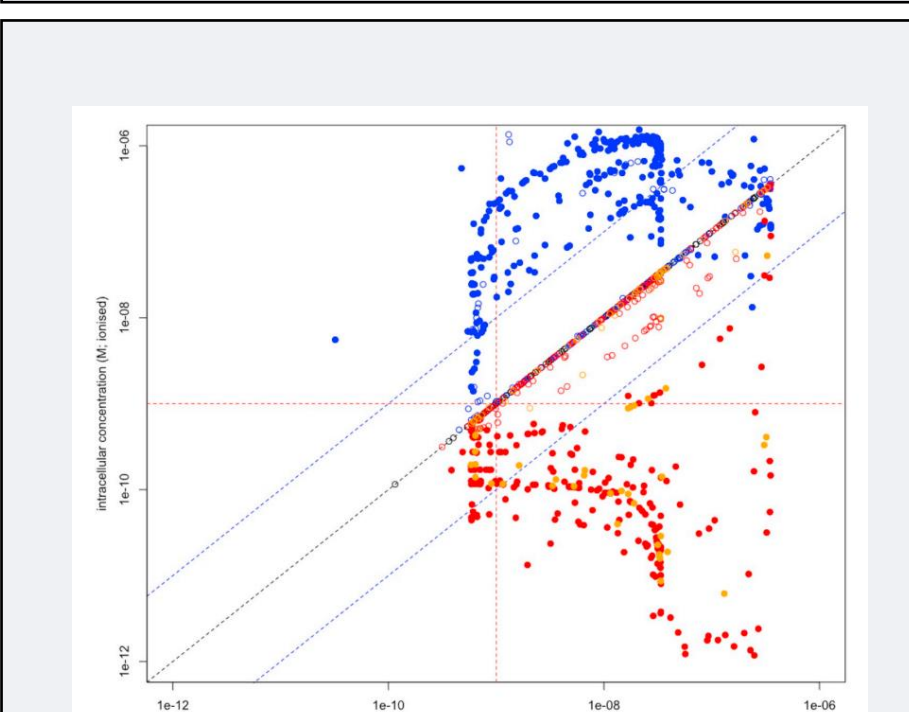
METHODS

A multi-pronged approach to determine the biologically effective concentration (BED) of MDCs.

1. Computational determination

2. Experimental determination in 2D and 3D *in vitro* liver models

Experimental plan and methods in 2D *in vitro* liver models



- 96-well plate design
- Usage of HepG2 cell line (human liver cancer cell line purchased from ATCC® HB-8065™)
- 48-h exposure with the respective MDC → conc.: 0.01; 0.1; 1; 10; 25 μM solution in 0.2 % DMSO
- Determination of C_{FREE}**
→ e.g., solid phase microextraction (SPME)
- Determination of C_{CELL}**
→ e.g., removing of medium followed by cell extraction with acetonitrile:water solution (1:2)
- Determination of C_{TOTAL}**
→ e.g., extraction from the exposure medium with acetonitrile

Computational modeling of C_{FREE} , C_{CELL} , C_{TOTAL} using the VIVD model [3]

Direct determination of C_{FREE} , C_{CELL} , C_{TOTAL}

- Determination of $C_{PLASTIC}$, $C_{EVAPORATED}$, C_{BOUND} for indirect determination of C_{FREE} and C_{CELL} and C_{TOTAL}**
→ e.g., DC assay (C_{BOUND}),...

$$C_{TOTAL} = C_{NOMINAL} - C_{PLASTIC} - C_{EVAPORATED} - C_{DEGRADATION}$$

$$C_{FREE} = C_{TOTAL} - C_{BOUND} - C_{CELL}$$

$$C_{CELL} = C_{TOTAL} - C_{BOUND} - C_{FREE}$$

- Measurements with HPLC-MS or GC-MS

Effective concentration *in vitro*

Indirect determination of C_{FREE} , C_{CELL} , C_{TOTAL}

This step will proceed in collaboration with Dr. Celline Brochet, the unit "Models for Ecotoxicology and Toxicology", INERIS, France.

Best approach

Quantitative *in vitro* to *in vivo* extrapolation (QIVIVE) i.e., physiologically based toxicokinetic modelling (PBTK) leads to:

Biologically effective concentration *in vivo*

PRELIMINARY RESULTS

- Experimental work:** The first analysis of BPA, BPF and BPS for direct determination of C_{TOTAL} and C_{CELL} showed high blind values (BV) → possible sources which led to contamination can be plastic lab equipment and contaminated water → first attempts were performed to discover sources to reduce BV:
 - Measurement of water blanks (2 water sources)
 - Usage of more glass ware and less plastic equipment
 - Method adaptation to reduce the contamination of BPA, BPF, BPS from the plastic
- Literature research:** Reviewing of available methods for the evaluation of C_{FREE} and C_{BOUND} .

OVERVIEW & PLANS FOR THE NEXT YEAR

- Usage of more sophisticated 3D *in vitro* models → for increased number of target sites; for better analyses of metabolism.
- Planned research stay abroad in the unit "Models for Ecotoxicology and Toxicology" INERIS (France). → Training to use *in vitro* kinetics models
- Poster presentations at the EUROTOX Congress 2021 and the 10th Annual Meeting of the ASCCT 2021



REFERENCES

[1] F. A. Groothuis, et al.: Dose metric considerations in *in vitro* assays to improve quantitative *in vitro*-*in vivo* dose extrapolation. *Toxicology* 332 (2015) 30-40

[2] R. Cano, et al.: Role of Endocrine-Disrupting Chemicals in the Pathogenesis of Non-Alcoholic Fatty Liver Disease: A Comprehensive Review. *International Journal of Molecular Sciences* 4807 (2021) 1-22

[3] C. Fisher, et al.: VIVD: virtual *in vitro* distribution model for the mechanistic prediction of intracellular concentrations of chemicals in *in vitro* toxicity assays. *Toxicology in Vitro* 58 (2019) 42-50