

# CYANOBACTERIAL TOXINS AND THE INTESTINES – WHAT ARE WE MISSING?

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## BACKGROUND

- Anthropogenic eutrophication of freshwater bodies and climate change increase the occurrence of toxic cyanobacterial **blooms**<sup>1</sup>
- Prolonged dry heat periods in summer pose a hazard to **drinking water safety**
- Upon (human) exposure to cyanotoxins, **gastrointestinal symptoms** (e.g. nausea, vomiting, diarrhea) are frequently reported<sup>2,3</sup>
- Most likely exposure route to water-borne cyanobacteria: accidental consumption of contaminated drinking water<sup>4</sup>
- Epithelia of the gastrointestinal tract (GIT) are the first barrier to be overcome for causing specific organ toxicities

• Most studied toxic effects (of isolated toxins)<sup>5</sup>:

- **Hepatotoxicity (1784)**
- **Neurotoxicity (1059)**
- **Carcinogenicity (929)**
- **Nephrotoxicity (729)**

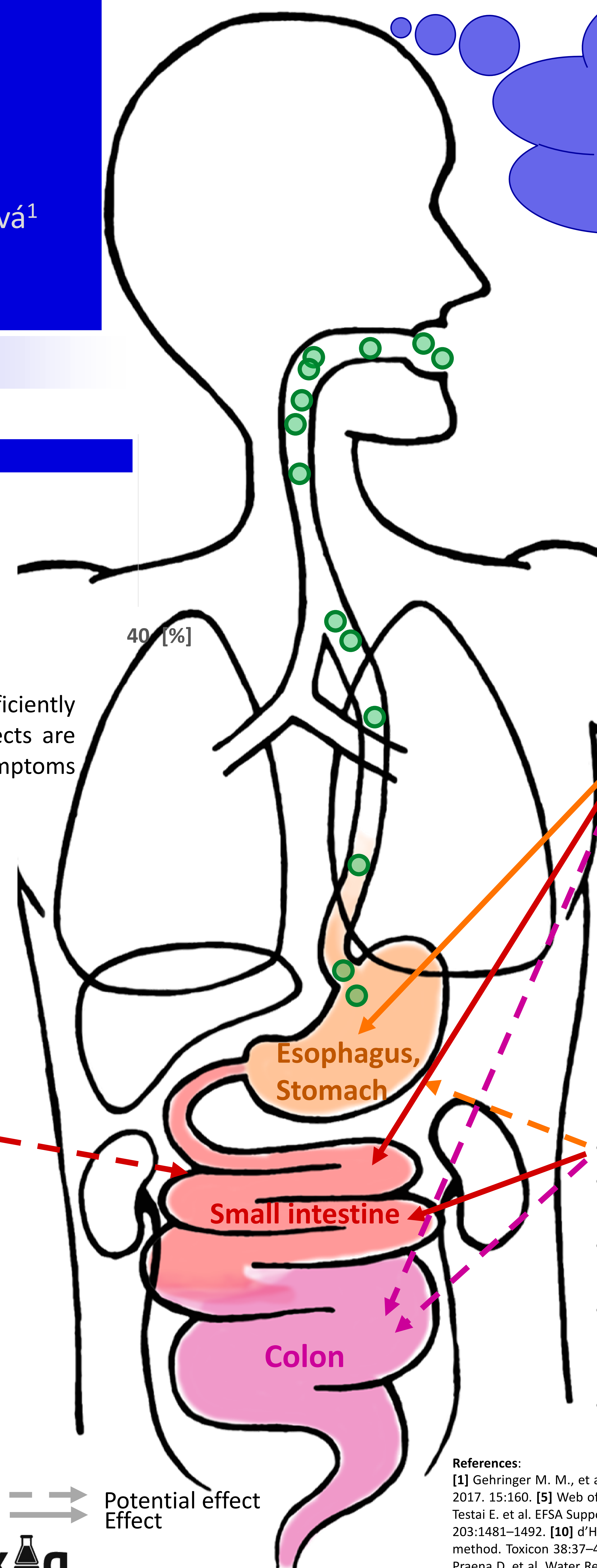
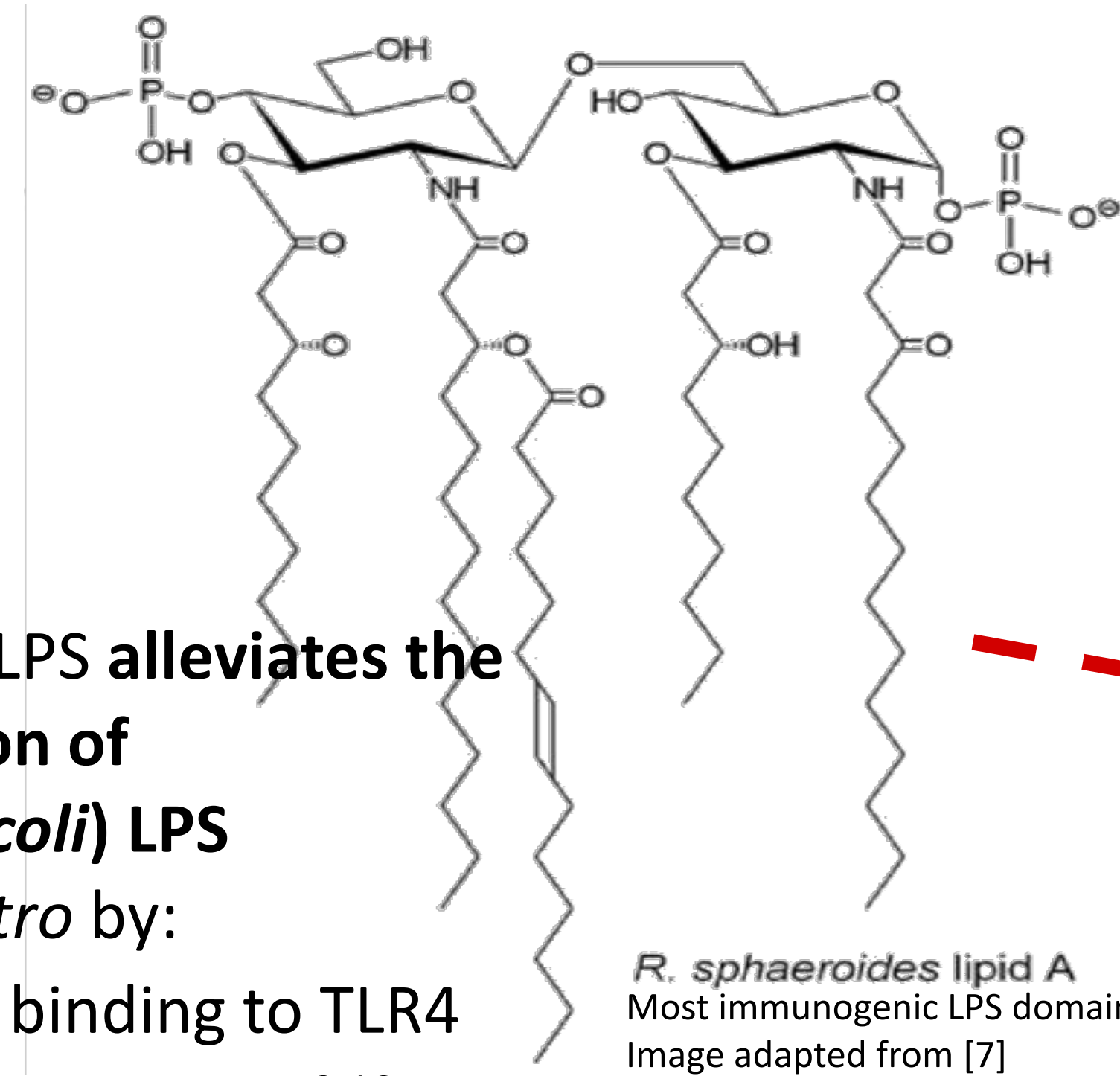
• Cyanobacterial metabolites diversity is not sufficiently reflected by toxicological studies, mixture effects are very likely to significantly contribute to the symptoms and illness severity<sup>6</sup>

## CYANOBACTERIAL LPS

- LPS = lipopolysaccharide
- Structural feature of the Gram-negative bacterial cell wall
- Recognized by the innate immune systems **pattern recognition receptor TLR4**, expressed particularly on epithelial, endothelial and immune cells (phagocytes)<sup>8</sup>
- Suggested to contribute to GIT inflammation
- Facilitate epithelial penetration of other cyanotoxins

**BUT:**

- Cyanobacterial LPS **alleviates the immune reaction of eubacterial (*E. coli*) LPS** *in vivo* and *in vitro* by:
- Competitively binding to TLR4
  - Potential for medical use<sup>9,10</sup>
  - Contribution to gastrointestinal irritation upon cyanobacterial intoxication is possible but yet **inconclusive** due to the lack of toxicological data
  - Cyanobacteria and their LPS structure are **phylogenetically distant** from eubacteria<sup>7</sup>



## OBJECTIVES

ADDRESSING THE DATA GAP: REVIEW OF FRESHWATER BLOOM EFFECTS ON THE GASTROINTESTINAL TRACT, INCLUDING:

- ENVIRONMENTAL MIXTURES
- **NOVEL CYANOBACTERIAL TOXINS & METABOLITES**
- IMPLICATIONS FOR **DRINKING WATER SAFETY**



## CONCLUSIONS & DATA GAPS

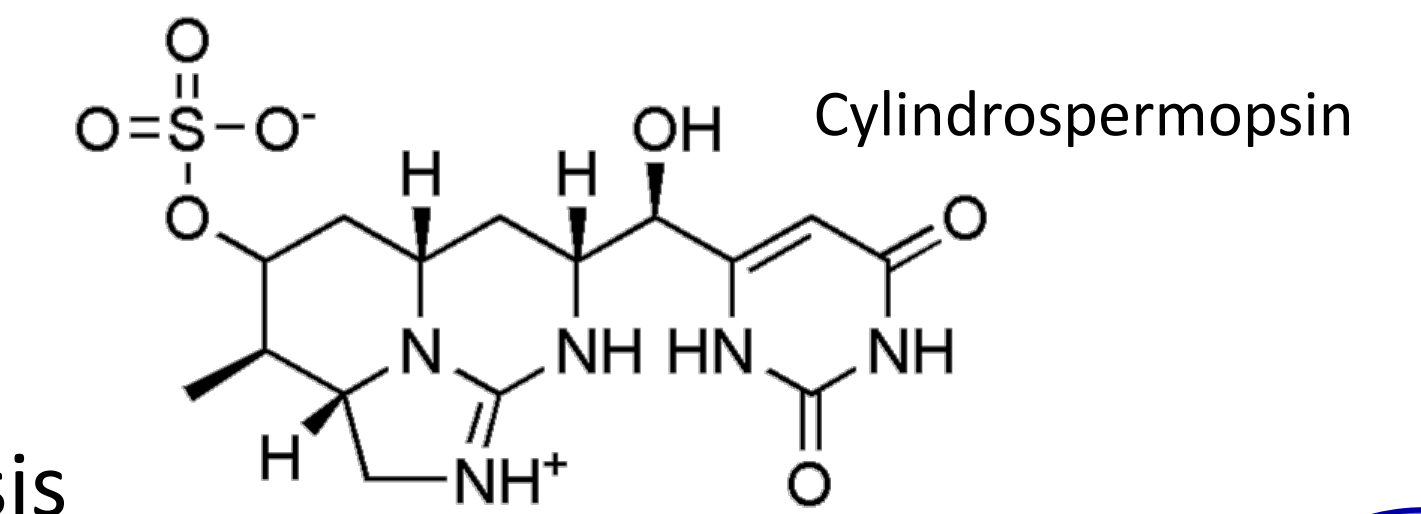
- Toxicity assessment of **novel toxins & metabolites** needed
- **Little information** on distinct gastro-intestinal effects
- Safe drinking Water: **Low toxin concentrations** need to be toxicologically covered
- **Mechanistic data** (e.g. from advanced *in vitro* assays) needed for accurate hazard characterization
- **Reassessment** of CYN and MC-LR for enterotoxicity recommended
- **Characterization** of the (non-)toxic **bloom metabolome**

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Effects of cyanobacterial toxins on the human gastrointestinal tract and the mucosal innate immune system  
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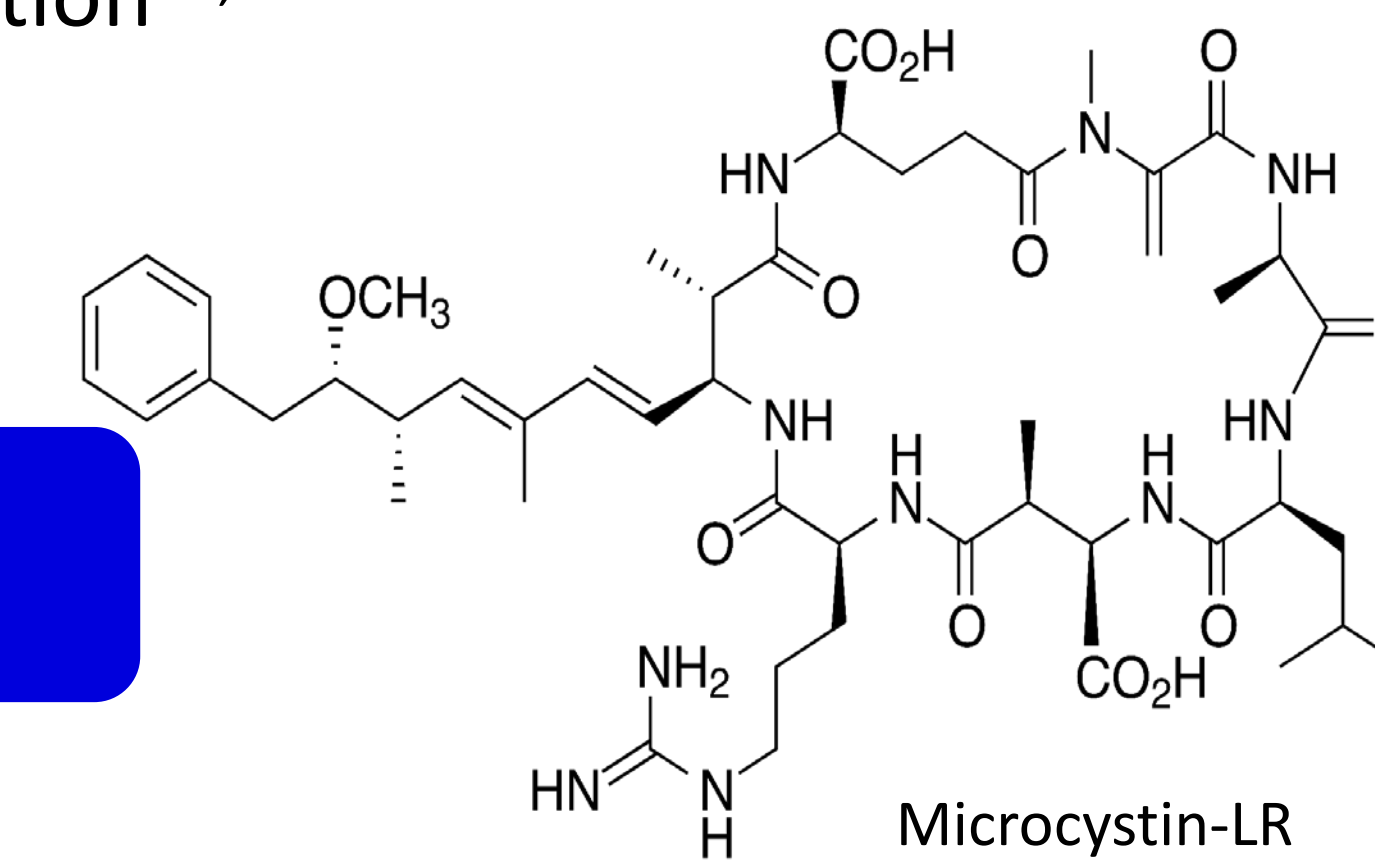
## CYLINDROSPERMOPSIN

- Hepatotoxin, cytotoxin
- Irreversible inhibition of protein biosynthesis
- Oral exposure causes ulceration of the **stomach** and lesions in the **(small) intestine** *in vivo* (mouse) and acts on **colon epithelia** *in vitro* (human, CaCo-2 cells)<sup>14,15,16</sup>
- **Pro-inflammatory** action<sup>17,18</sup>



## MICROCYSTINS

- Hepatotoxins
- Irreversible inhibitors of the ubiquitous intracellular protein phosphatase 1 and 2A
- **Uptake** via organic anion transport polypeptides and the bile acid system in the **small intestine**
- Besides liver, impact also the **(small) intestine** upon *in vivo* oral exposure (rodent models) and **colon epithelia** *in vitro* (human, CaCo-2 cells)<sup>11,12,13</sup>
- Activate macrophages *in vitro*<sup>14</sup>



WHAT ABOUT OTHER COMPOUNDS?

## MIXTURES

- Blooms: biomass + exudate
- **Most probable** form of exposure to cyanobacteria
- **Highly variable** in biological and chemical composition (associated bacteria, cyanobacterial taxa, metabolites...), **poorly characterized**<sup>6</sup>
- Symptoms upon exposure: **gastroenteric disease, nausea, diarrhoea, abdominal pain**<sup>3,19</sup>
- → **co-action** of many factors

References:

[1] Gehring M. M., et al. Int J Biochem & Cell Biol, 2004. 36(5):931-941. [2] Svirčev Z. et al. Arch Tox 2017. 91:621–65. [3] Levesque B. et al. Sci Tot Env 2014. 466–467:397–403. [4] Miller T.R. et al. Mar Drugs 2017. 15:160. [5] Web of Knowledge search tags (2018-10-15, 12:42 CEST): TS=(cyano\* AND hepatotox\*), TS=(cyano\* AND (nephrotox\* OR kidney\*)), TS=(cyano\* AND neurotox\*), TS=(cyano\* AND carcino\*) [6] Testai E. et al. EFSA Support Publ 13, 2016. doi: 10.2903/SP.EFSA.2016.EN-998 [7] Stewart I. et al. Environ Health, 2006. 5:7. [8] Vaure C. and Liu Y. Front Immun, 2014. 5:316. [9] Macagno A. et al. J Exp Med, 2006. 203:1481–1492. [10] d’Hennezel E. et al. mSystems, 2017. 2. [11] Ito E, Kondo F, Harada K-I (2000) First report on the distribution of orally administered microcystin-LR in mouse tissue using an immunostaining method. Toxicon 38:37–48. [12] Botha N. et al. Toxicon, 2004. 43:251–254. [13] Fastner J. et al. Toxicon, 2003. 42:313–321. [14] Adamovsky O. et al. Environ Sci Technol, 2015. 49:12457–12464. [15] Gutiérrez-Praena D. et al. Water Res, 2012. 46:1566–1575. [16] Pichardo S. et al. Toxins, 2017. 9:402. [17] Sierosławska A. et al. J Appl Toxicol, 2015. 35:1406–1414. [18] Poniedziałek B. et al. Chemosphere, 2015. 120:608–614. [19] Svirčev Z. et al. Arch Toxicol, 2017. 91:621–650. [20]

Potential effect  
Effect



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